



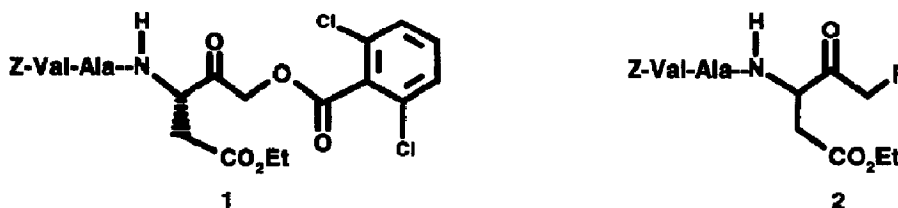
## Synthesis of P1 Aspartate-Based Peptide Acyloxymethyl and Fluoromethyl Ketones as Inhibitors of Interleukin-1 $\beta$ -Converting Enzyme

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**Abstract:** Improved procedures have been developed for the synthesis of P1 aspartate-based 2,6-dichlorobenzoyloxymethyl ketone **1** and fluoromethyl ketone **2**, the prodrugs of two potent ICE-inhibitors. **1** was prepared from (*R*)-*trans*-4,5-*O*-isopropylidene-4,5-dihydroxy-2-pentenecarboxylic acid ethyl ester; **2** was obtained via a nitro-aldol condensation as key step from *in situ* generated fluoroacetaldehyde.

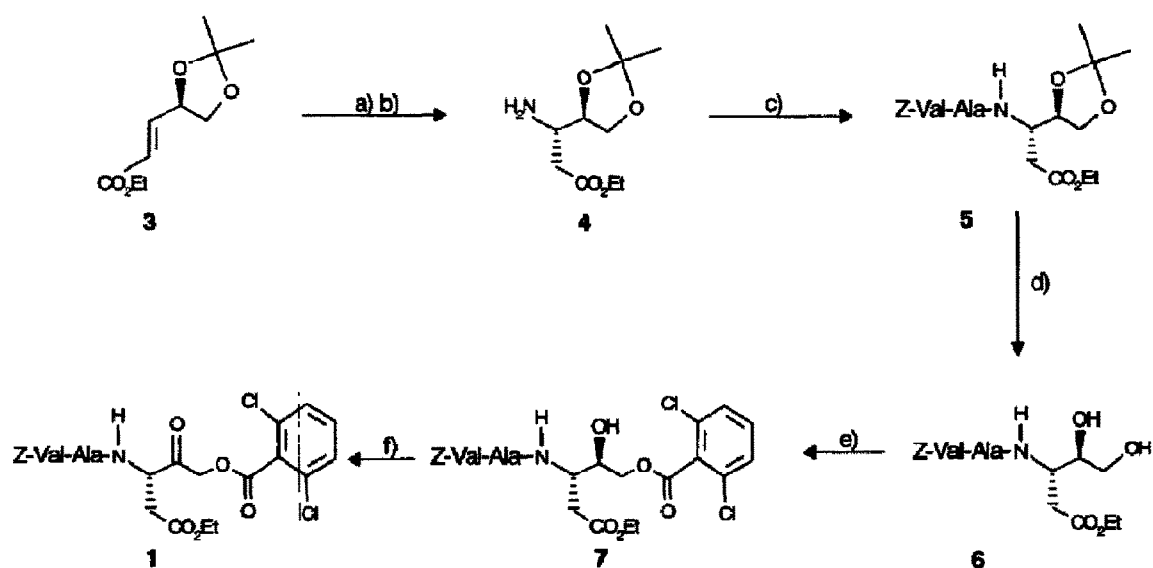
ICE (Interleukin-1 $\beta$ -converting enzyme) is a recently discovered atypical cysteine protease, which processes an inactive precursor to the proinflammatory cytokine IL-1 $\beta$  and may regulate programmed cell death in neuronal cells [1, 2]. Since specific inhibitors of ICE may be of benefit in inflammatory and degenerative neuronal diseases such as Rheumatoid Arthritis, Alzheimer's and Parkinson's diseases, we wanted to test their *in vivo* potential in animal models and needed large amounts of **1** and **2**, the ethyl ester prodrugs of two potent ICE-inhibitors [3,4].



Current methods for preparing acyloxymethyl ketones as thiol protease inhibitors use diazomethane [5] for the homologation of a conveniently protected amino acid, where the resulting diazomethyl ketone is converted to the corresponding bromide and then reacted with an acid to provide the desired product. We have developed a method suitable for the large scale preparation of aspartate-based acyloxymethyl ketones of type

1, avoiding the use of diazomethane (*Scheme 1*). Commercially available (*R*)-*trans*-4,5-*O*-isopropylidene-4,5-dihydroxy-2-pentenecarboxylic acid ethyl ester **3** was treated with benzylamine at  $-20^{\circ}\text{C}$  for 40 h according to [6] and rendered the desired addition product in 75% yield, which was hydrogenated to the amine **4** and isolated as crystalline oxalate.

*Scheme 1*



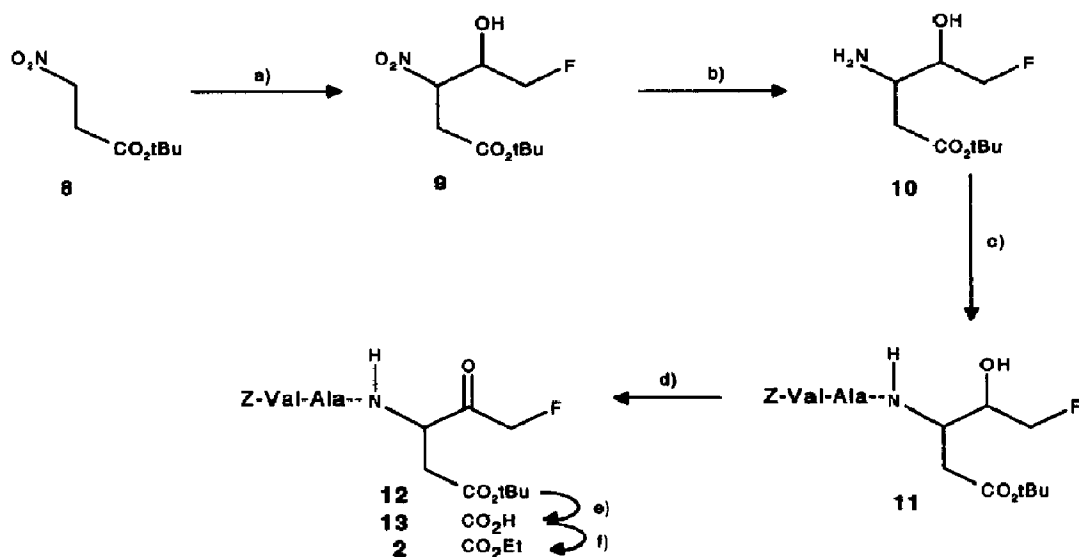
a) Benzylamine,  $-20^{\circ}\text{C}$ , 40hrs. Chromatography, 75% b)  $\text{H}_2$ , Pd/C, EtOH. Oxalate crystallized from EtOAc/Ether, 95%.  
 c) Z-Val-Ala-OH, EDCI, HOBT, THF,  $0^{\circ}\text{C}$  to r.t., 12 hrs., 66%. d) EtOH/H<sub>2</sub>O (5:1) K10 Montmorillonite, 2.75hrs,  $75^{\circ}\text{C}$ , 73%. e) 2,6-Dichlorobenzoylchloride, pyridine, DMAP (2eq.), DMPU (2eq.), 2 days, 93%. f) Dess-Martin,  $0^{\circ}\text{C}$  to r.t. 3.5hrs, 90%.

The free base of **4** was coupled to Z-Val-Ala-OH under standard conditions (HOBT, EDCI, THF) to yield ketal **5**, which was selectively hydrolyzed to diol **6** with Montmorillonite K10 in EtOH/water. The latter was esterified with 2,6-dichlorobenzoyl chloride in pyridine/DMAP using DMPU as cosolvent. Several methods were elaborated for the final oxidation step, since ketone **7** was sensitive to epimerization under basic conditions. Swern oxidation e.g. gave a ~1:1 mixture of epimers, while the procedures of Dess-Martin and Pfitzner-Moffatt

[8] gave pure 1.

Fluoromethyl ketones are generally prepared with limited success according to a modified Dakin West procedure [9], which has been used by R. Black in [4b] for the synthesis of BOC-Asp-CH<sub>2</sub>-F. We decided to synthesize 2 via nitro-aldol condensation [10] as a key step (*Scheme 2*). For this purpose, fluoroethanol was submitted to a Swern oxidation, which was followed by the addition of the nitro derivative 8 without isolation of the intermediate fluoroacetaldehyde. The yield of nitroalcohol 9 resulting from this one-pot oxidation/condensation reaction amounted to 89% after purification by chromatography. Hydrogenation over Raney-Nickel gave amino alcohol 10 in 85% yield, which was converted to 11 by coupling with Z-Val-Ala-OH under standard conditions. Dess-Martin oxidation proceeded in 62% yield to the keto ester 12 as a crystalline solid. Deprotection of the *tert*.butyl ester with TFA provided the acid 13, which was esterified in HCl/EtOH over night at r.t. to generate 2, obtained as a crystalline solid and according to NMR as a -1:1 mixture of epimers [7]. No attempt was made to separate the epimers.

**Scheme 2**



a) 2-Fluoroethanol, (COCl)<sub>2</sub>, DMSO, NEt<sub>3</sub>, -60°C to r.t., 1.5 hrs, then add 8, r.t. 1hr. 89%. b) Raney-Nickel, MeOH, H<sub>2</sub>, r.t. 12 hrs. 85%. c) Z-Val-Ala-OH, EDCI, HOBT, DMAP, THF, 12 hrs. 97% white solid. d) Dess-Martin reagent (7eq.), CH<sub>2</sub>Cl<sub>2</sub>, 30min. 62% white solid. e) TFA, CH<sub>2</sub>Cl<sub>2</sub>, 30 min. r.t. 91% white solid. f) EtOH saturated with HCl, 14 hrs. r.t. 86% white solid.

In models of acute inflammation, such as the LPS-pyrexia and Carrageenan oedema, **1** and **2** showed potent effects upon oral administration with the ED<sub>50</sub>'s less than 0.1 mg/kg p.o. Whether these activities are mediated by inhibition of IL-1 $\beta$ , is under current investigation.

#### Acknowledgements:

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#### References and Notes:

- [1] Wilson, K.P.; Black, J.F.; Thomson, J.A.; Kim, E.E.; Griffith, J.P.; Navia, M.A.; Murcko, M.A.; Chambers, S.P.; Aldape, R.A.; Raybuck, S.A. and Livingston, D.J. *Nature*, **1994**, *370*, 270.
- [2] Walker, N.P.C.; Talanian, R.V.; Brady, K.D.; Dang, L.C.; Bump, N.J.; Ferenz, C.R.; Franklin, S.; Ghayur, T.; Hackett, M.C.; Hammill, L.D.; Herzog, L.; Hugunin, M.; Houy, W.; Mankovich, J.A.; McGuinness, L.; Orlewicz, E.; Paskind, M.; Pratt, C.A.; Reis, P.; Summani, A.; Terranova, M.; Welch, J.P.; Xiong, L.; Moeller, A.; Tracey, D.E.; Kamen, R. and Wong, W.W. *Cell*, **1994**, *78*, 343.
- [3] Dolle, R.E.; Hoyer, D.; Prasad, C.V.C.; Schmidt, S.J.; Helaszek, C.T.; Miller, R.E. and Ator, M.A. *J.Med.Chem.* **1994**, *37*, 563.
- [4] a) Heng, R.; Payne, T.G.; Revesz, L. and Weidmann, B. *PCT Application*, WO 93/09135. *Chem.Abstr.* **1994**, *120*, 77637.  
b) Black, R.; Sleath, P.R. and Kronheim, S.R. *PCT Application*, WO 91/15577. *Chem.Abstr.* **1992**, *116*, 79400.
- [5] Smith, R.A.; Copp, L.J.; Coles, P.J.; Pauls, H.W.; Robinson, V.J.; Spencer, R.W.; Heard, S.B. and Krantz, A. *J.Am.Chem.Soc.*, **1988**, *110*, 4429.
- [6] Matsunaga, H.; Sakamaki, T.; Nagaoka, H. and Yamada, Y. *Tetrahedron Lett.* **1983**, *24*, 3009.
- [7] Diagnostic physical data for **1**: M.p.: 193<sup>o</sup> C; [ $\alpha$ ]<sub>D</sub><sup>20</sup> = -52.0<sup>o</sup> C [c=1.05 in acetone]. <sup>13</sup>C NMR (360MHz; DMSO)  $\delta$  13.98; 17.71; 18.07; 19.21; 30.33; 34.25; 48.10; 52.88; 59.86; 60.34; 65.40; 67.72; 127.58; 127.69; 128.26; 128.44; 130.79; 132.51; 137.00; 156.10; 163.20; 170.09; 170.98; 172.90; 199.44.  
For **2**: <sup>1</sup>H NMR (360MHz, DMSO-d<sub>6</sub>, 120<sup>o</sup>C)  $\delta$  0.84-0.92(m,6H); 1.18 (t, 3H, J=8Hz); 1.22 and 1.25 (two d, 3H, J=8Hz, Me of Ala); 1.96-2.09 (m, 1H); 2.58-2.67 (dd, 1H); 2.80-2.88 (m, 1H); 3.88-3.93 (m, 1H); 4.08(q, 2H, J=8Hz); 4.23-4.35(m, 1H); 4.60-4.72(m, 1H); 4.95-5.23(m, 2H, CH<sub>2</sub>F); 5.05 (d, 2H, PhCH<sub>2</sub>O); 6.62(bd, 1H, J=9Hz); 7.25-7.33(m, 5H); 7.60-7.70(bq, 1H); 7.98-8.05(bt, 1H)
- [8] Ireland, R.E. and Liu, L. *J.Org.Chem.* **1993**, *58*, 2899. Pfitzner, K.E. and Moffatt, J.G. *J.Am.Chem.Soc.* **1963**, *85*, 3027.
- [9] Rasnick, D. *Anal.Biochem.* **1965**, *149*, 461.
- [10] Imperiali, B.; Abeles, R.H. *Tetrahedron Lett.* **1986**, *27*, 135.

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